



## Instrumentation and Research

**Table 1.** BT and GU research projects at CLS-SM in 2009

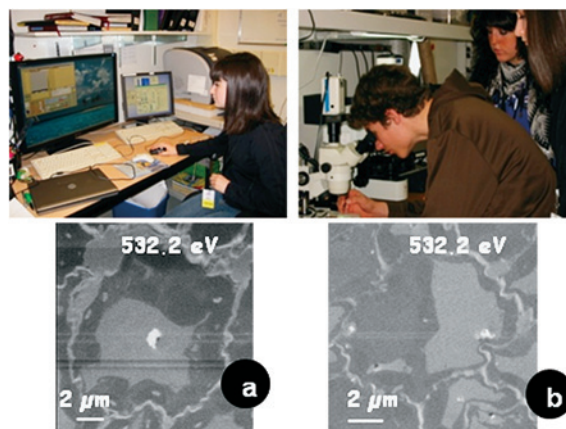
PI	Title
<b>Materials Science</b>	
A.P. Hitchcock (McMaster)	X-ray magnetic circular dichroism of individual magnetosomes
A. P. Chandra (South Australia)	A PEEM and NEXAFS study of acid leached pyrite surface
B. van Waeyenberge (Max Planck Institute)	Fast vortex core dynamics in magnetic nano structures
F. De Groot (Utrecht)	In-situ X-ray spectromicroscopy of iron-oxide Fischer-Tropsch catalysts
M. Pézolet (Laval)	Mapping orientation in silk fibres by STXM
S.G. Urquhart (Saskatchewan)	XPEEM studies of phase segregation in Langmuir-Blodgett films; TEY-STXM
V. Coker (Manchester)	STXM analysis of bio-nanomagnetic spinels
Y. Mozharivskiy (McMaster)	XMCD of magnetic semiconductors for spintronics
<b>Environmental Science</b>	
A. Prange (Bonn)	Speciation of sulphur globules in bacterial cells at the S 2p edge
B. De Gregorio (U.S. Naval Research Lab)	Stardust and meteorites: studies of Primitive Extraterrestrial Organic Matter
C. Rivard (France)	Iron speciation in clay minerals under anoxic conditions
J. Lawrence (NWRI)	Mapping antimicrobial agents and their effects on bacterial biofilms
J. Lehmann (Cornell)	Mineral control over carbon sequestration in soil
K. Norlund (McMaster)	Microbial sulphur metabolism in acid mine drainage
M. Obst (Tuebingen)	Sorption of phenanthren to black carbon particles
I. Pickering (Saskatchewan)	Selenium speciation in bacterial biofilms
<b>Life Science</b>	
C. Gaillard (INRA)	Correlative microscopy of biopolymers in seeds
D. Korber (Saskatchewan)	Effect of anti microbial agents on wild and mutant type <i>Escherichia coli</i>
J. Gardella (SUNY Buffalo)	Spatial and chemical mapping of a biodegradable polymer thin films
J. Wang (CLSI)	3D chemical and elemental imaging of biological samples by STXM tomography
K. Kaznatcheev (BNL)	Extended soft x-ray dichroic measurements of large molecules
M. Ryan (Manchester)	Compatibility of nanomaterials in human tissue near hip replacements
S. Turgeon (Laval)	Plasma Deposited Fluoropolymer Coatings for Stent Applications

The development of instrumentation and techniques for humid cell, wet cell, tomography, and total electron yield (TEY) in STXM are actively pursued in collaborations of beamline staff with beam team members. Soft X-ray tomography is able to generate 3D chemical mapping and localized information on materials, biological and environmental samples [12]. The development of a

controlled humidity cell and a novel wet cell enhances the capability to study the effect of humidity on different samples and to study biological samples in their native forms [13]. The TEY capability with STXM will enable us to determine the surface and bulk properties of samples simultaneously while exploiting the high spatial resolution of the STXM [14].

## Outreach Activities

The beam team and beamline staff members actively participate in outreach activities each year. During 2009, Grade 11 and 12 students from Avonlea School in Saskatchewan used the beamline to study the “Effect of UV light on Plant Cells” by exposing canola plants to UV light and determined the structural and chemical changes to canola leaf cells using STXM.



**Figure 2:** (Top row) Avonlea School students learning to use optical and STXM microscopes at the beamline; (bottom) X-ray images of UV exposed (a) and unexposed (b) canola leaf cells recorded at the O 1s edge showing differences in the morphology and composition.

## References

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